Anti-ulcerogenic activity of stem extract and fractions of Homalium letestui

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ABSTRACT

Background & Aim: Homalium letestui Pellegr (Flacourtiaceae) used traditionally by the Ibibios of Southern Nigeria to treat stomach ulcer, malaria and other inflammatory diseases was evaluated for antiulcer properties.

Experimental: The effects of ethanol stem extract of H. letestui (200 – 600 mg/kg) and fractions (Aqueous and Dichloromethane, 400 mg/kg) on experimentally induced ulcer were studied in rats using ethanol, indomethacin, reserpine and histamine –induced ulcer models.

Results & Discussion: The extract (200 – 600 mg/kg) inhibited ethanol, indomethacin, reserpine and histamine –induced ulcer models in a dose dependent fashion. The various degrees of inhibitions were statistically significant (p<0.05, 0.01, 0.001). The effects of the extract/fraction were comparable to that of the standard drugs used in indomethacin and ethanol-induced ulcer models with the dichloromethane fraction having the highest activity.

Industrial and practical recommendations: The present study demonstrates that stem extract of Homalium letestui might be useful for the treatment of ulcer.

1. Introduction

Homalium letestui Pellegr (Flacourtiaceae) is a forest tree growing up to 80–100 ft and found in the rainforest of West Africa (Hutchinson and Daziel, 1963; Keay, 1989). The plant parts; particularly stem bark and root are used in various decoctions traditionally by the Ibibios of the Niger Delta of Nigeria to treat stomach ulcer, malaria and other inflammatory diseases as well as an aphrodisiac (Okokon et al., 2006). Reports of antiplasmodial (Okokon et al., 2006), antidiabetic (Okokon et al., 2007), anti-inflammatory and analgesic (Okokon et al., 2013a), cellular antioxidant, anticancer, and antileishmanial (Okokon et al., 2013b), depressant and anticonvulsant (Okokon and Davies, 2014)
activities of the plant have been published. We report in this study the antulcerogenic activity of this plant in order to provide scientific basis for its use in traditional medicine in the treatment of stomach ulcer.

2. Materials and Methods

2.1. Plants collection

The plant material *Homalion letestui* (stem) was collected in a forest in Uruan area, Akwa Ibom State, Nigeria in July, 2014. The plant was identified and authenticated by Dr. Margaret Bassey of Department of Botany and Ecological Studies, University of Uyo, Uyo, Nigeria. Herbarium specimen (FPUU 382) was deposited at Department of Pharmacognosy and Natural Medicine Herbarium.

2.2. Extraction

The stem was washed and shade-dried for two weeks. The dried plant material was further chopped into small pieces and reduced to powder. The powdered material was macerated in 70% ethanol. The liquid filtrates were concentrated and evaporated to dryness in vacuo at 40°C using rotary evaporator. The crude ethanol extract (10 g) was partitioned with a 50:50 mixture of distilled water and dichloromethane. The aqueous fraction was evaporated to dryness in a water bath at 60°C and the dichloromethane fraction air-dried. The ethanol extract, the aqueous and dichloromethane fractions were stored at -4°C until used.

2.3. Animals

Albino wistar rats (168 – 175 g) of either sex were obtained from the University of Uyo animal house. They were maintained on standard animal pellets and water *ad libitum*. Permission and approval for animal studies were obtained from the College of Health Sciences Animal Ethics committee, University of Uyo.

2.4. Indomethacin induced ulcer

Male adult albino rats were used for the experiment. They were randomized into seven groups of six rats each. Food was withdrawn 24 hours and water 2h before the commencement of experiment (Alphin and Ward, 1967). Group 1(control) received only ethanol (2.5 ml/kg p.o), Groups 2-4 were pretreated with *H. letestui* stem extract (200, 400 and 600 mg/kg p.o. respectively); Group 5 received aqueous fraction (400 mg/kg); Group 6 received dichloromethane fraction (400 mg/kg), and Group 7, cimetidine (100 mg/kg p.o. dissolved in 5% Tween 80). One hour later, groups 2 - 7 were administered with indomethacin. Four hour after indomethacin administration, animals were killed by cervical dislocation. The stomachs were removed and opened along the greater curvature. The tissues were fixed with 10% formaldehyde in saline. Macroscopic examination was carried out with a hand lens and the presence of ulcer lesion was scored (Nwafor et al., 1996). Ulcer index (UI), preventive ratio (PR) and degree of ulceration (DU) of each of the groups pretreated with extract were calculated using standard methods (Zaidi and Mukerji 1958; Nwafor et al., 2000).

2.5. Ethanol induced gastric ulceration

The procedure was similar to that used in indomethacin induced ulceration. The rats randomly assigned into were randomized into eight groups of six rats each. Food was withdrawn 24 hours and water 2h before the commencement of experiment (Alphin and Ward, 1967). Group 1(control) received only ethanol (2.5 ml/kg p.o), Groups 2-4 were pretreated with *H. letestui* stem extract (200, 400 and 600 mg/kg p.o. respectively); Group 5 received aqueous fraction (400 mg/kg); Group 6 received dichloromethane fraction (400 mg/kg) and Group 7, received propranolol (40 mg/kg p.o. dissolved in distilled water). One hour later, groups 2 -6 were administered with ethanol. Four hour after ethanol administration, animals were killed by cervical dislocation. The stomachs were removed and opened along the greater curvature. The tissues were fixed with 10% formaldehyde in saline. Macroscopic examination was carried out with a hand lens and the presence of ulcer lesion was scored (Nwafor et al., 2000).

2.6. Histamine-induced gastric ulceration in rats

The procedures were similar to that used in indomethacin-induced ulceration except that the negative control group (group 1) received only histamine acid phosphate (Sigma, 100 mg/kg i.p. dissolved in distilled water) (Maity et al., 1995). Groups 2 - 4 were pretreated with *H. letestui* extract (200, 400 and 600 mg/kg p.o. respectively); Group 5 received aqueous fraction (400 mg/kg), Group 6 received dichloromethane fraction (400 mg/kg) and
Group 7 received cimetidine (100 mg/kg p.o. dissolved in 50% Tween 80). One hour later, groups 2 - 7 were administered with histamine acid phosphate, 100 mg/kg i.p. Eighteen (18) hours after histamine administration, animals were killed by cervical dislocation. The stomachs were removed and opened along the greater curvature. The tissues were fixed with 10% formaldehyde in saline. Macroscopic examination was carried out with a hand lens and the presence of ulcer lesion was scored (Nwafor et al., 1996), stomach processing and examination as well as ulcer scoring were similar to that used in indomethacin-induced ulceration.

2.7. Reserpine induced gastric ulceration in rats

Male adult albino rats weighing 120 – 170 g were used for the experiment. They were randomized into six groups of six rats each. Food was withdrawn 24 hours and water 2 h before the commencement of experiment (Alphin and Ward, 1967). Group 1 (control) received only reserpine (Sigma, 8 mg/kg p.o. dissolved in Tween 80); Groups 2 - 4 were pretreated with H. letestui extract (200, 400 and 600 mg/kg p.o. respectively); Group 5 received aqueous fraction (400 mg/kg). Group 6 received dichloromethane fraction (400 mg/kg) and Group 7 received cimetidine (100 mg/kg p.o. dissolved in 50% Tween 80) (Maity et al., 1995). Eighteen hours (18 h) after reserpine administration, animals were killed by cervical dislocation. The stomachs were removed and opened along the greater curvature. The tissues were fixed with 10% formaldehyde in saline. Macroscopic examination was carried out with a hand lens and the presence of ulcer lesion was scored (Nwafor et al., 1996). Ulcer index (UI), preventive ratio (PR) and degree of ulceration (DU) of each of the groups pretreated with extract were calculated using standard methods (Zaidi and Mukerji, 1985; Nwafor et al., 2000).

2.8. Statistical analysis

Data are reported as mean ± standard error of the mean (SEM) and were analyzed statistically using One way ANOVA followed by Tukey-kramer multiple comparison test and values of p<0.01 were considered significant.

3. Results and discussions

3.1. Indomethacin induced gastric ulceration

The extract and fractions (p.o.) pretreatment on indomethacin induced gastric ulceration showed a dose dependent reduction in ulcer indices in pretreated groups relative to control. The reductions were statistically significant (p<0.05, 0.001) compared to control. The dichloromethane fraction exerted the highest antiulcerogenic effects which was comparable to that of the standard drug used, cimetidine (Table 1).

3.1. Ethanol induced gastric ulceration

The extract and fractions significantly protected rats from ethanol – induced ulcer (Table 2). There was a significant (p<0.001) dose-dependent reduction in the ulcer indices relative to control. The effect of the extract and fractions were less than that of the standard drug, propranolol.

3.2. Histamine – induced ulceration

Administration of the extract and fractions significantly (p<0.001) reduced histamine-induced gastric ulceration in a dose dependent fashion compared to control (Table 3). The dichloromethane exhibited a higher antiulcer potential than the aqueous fraction but less than that of the standard drug cimetidine.

3.3. Reserpine – induced ulceration

Administration of the extract and fractions significantly (p<0.001) reduced reserpine induced gastric ulceration in a dose dependent fashion compared to control (Table 3). These effects were incomparable to that of the standard drug, cimetidine.

Homalium letestui stem has been reported to be used traditionally in the treatment of ulcer (Okokon et al., 2006). For this reason, the antiulcer activity of the stem bark extract and fractions was evaluated using indomethacin, ethanol, reserpine and histamine–induced ulcer models. Indomethacin, a known ulcerogen especially in an empty stomach (Bhargava et al., 1973) causes ulcer mostly on the glandular (mucosal) part of the stomach (Evbuonwa and Bolarinwa, 1990; Nwafor et al., 1996) by inhibiting prostaglandin synthetase through the cyclooxygenase pathway (Rainsford, 1987). Prostaglandins function to protect the stomach from injury by stimulating the secretion of bicarbonate and mucus, maintaining mucosal blood flow and regulating mucosal turn over.
and repair (Hayllar and Bjarnason, 1995; Hiruma-Lima et al., 2006).

Suppression of prostaglandin synthesis by indomethacin results in increased susceptibility of stomach to mucosal injury and gastroduodenal ulceration. The extract was observed to significantly reduce mucosal damage in the indomethacin – induced ulcer model, suggesting the possible extract mobilization and involvement of prostaglandin in the anti-ulcer effect of the extract. Administration of ethanol has been reported to cause disturbances in gastric secretion, damage to the mucosa, alterations in the permeability, gastric mucus depletion and free radical production (Salim, 1990). This is attributed to the release of superoxide anion and hydroperoxy free radicals during metabolism of ethanol as oxygen derived free radicals has been found to be involved in the mechanism of acute and chronic ulceration in the gastric mucosa (Pihan et al., 1987). It was observed in this study that the extract reduced significantly ethanol-induced ulcer. This may be due to cytoprotective effect of the extract via antioxidant effects which the stem extract has been reported to exhibit (Okokon et al., 2013b).

Okokon et al., (2013a) reported the presence of α-terpineol, Vanillin, 4-phenyl isocoumarin, 3,4,5-trimethoxy phenol, 2-Coumaranone, and xanthones in the stembank extract of H. letestui. Vanillin, a phenolic aldehyde has been reported to possess antioxidant and free radical scavenging ability (Kamat et al., 2000; Kumar et al., 2002; Lirdprapamongkol et al., 2009) which could possibly account for the anti-ulcer property of this plant.

α-terpineol present in this extract, is an isomer of the monoterpene, terpinen-4-ol. α-terpineol and terpinen-4-ol have been reported to possess anti-ulcer activity (Matsunaga et al., 2000; Souza et al., 2011). The antiulcer activity of this extract could also be due to the present of α-terpineol. Similarly, xanthones have been reported to demonstrate antiulcer activity (Ali et al., 2014). These compounds present in the stem extract maybe responsible for the observed antiulcer activity.

Ethanol is also reported to cause gastric mucosal damage by stimulating the formation of leukotriene C4 (LTC4) (Whittle et al., 1985). The gastroprotective effect of the extract may in part be due to the suppression, by the extract of lipoxygenase activity (Nwafor et al., 1996). Histamine-induced ulceration is known to be mediated by enhanced gastric acid secretion as well as by vasospastic action of histamine (Cho and Pfeiffer, 1981). The inhibition of ulcer due to histamine by the extract may be due to its suppression of histamine-induced vasospastic effect and gastric secretion.

Okokon et al., (2007) had reported the presence of flavonoids, saponins, terpenes and tannins in the stem extract of H. letestui. Flavonoids such as quercetin has been reported to prevent gastric mucosal lesions in various experimental models (Di carlo et al., 1999; Zayachkivska, 2005) by increasing the amount of neutral glycoproteins (Di carlo et al., 1999). Flavonoids have been reported to protect the gastric mucosa from damage by increasing the mucosal prostaglandin content and by inhibiting histamine secretion from mast cells by inhibition of histidine decarboxylase. Free radical scavenging ability of flavonoids has been reported to protect the gastrointestinal tract from ulcerative and erosion lesion (Borreli and Izzo, 2000). Saponins, especially triterpenes type have been implicated in antiulcer activity mediated by formation of protective mucus on the gastric mucosa and also protect the mucosa from acid effects by selectively inhibiting PGF2α (Agwu and Okunji, 1986; Lewis and Hanson, 1991).

4. Conclusions

The results of the present study show that stem extract and fractions of H. letestui displays gastroprotective activity as demonstrated by significant inhibition of the formation of ulcers induced through four different ulcer models studied. The antiulcer activity of the extract maybe due to the action of its phytochemical compounds present in the extract. The observation justifies the ethnomedical uses of the plants as antiulcer and antacid in addition to its nutritional values.

5. Acknowledgements

The authors are grateful to Ms. Sifon Akpan of Pharmacology and Toxicology Dept for her technical assistance.
Table 1. Effect of *Homalium letestui* extract on indomethacin induced ulcer.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Dose (mg/kg)</th>
<th>Ulcer indices</th>
<th>Preventive ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (indomethacin)</td>
<td>60</td>
<td>18.34 ± 2.72</td>
<td>-</td>
</tr>
<tr>
<td><em>Homalium letestui</em> extract p.o.</td>
<td>200</td>
<td>10.05 ± 1.22&lt;sup&gt;a&lt;/sup&gt;</td>
<td>45.20</td>
</tr>
<tr>
<td></td>
<td>400</td>
<td>2.10 ± 0.60&lt;sup&gt;c&lt;/sup&gt;</td>
<td>88.54</td>
</tr>
<tr>
<td></td>
<td>600</td>
<td>2.00 ± 0.76&lt;sup&gt;c&lt;/sup&gt;</td>
<td>89.09</td>
</tr>
<tr>
<td>Dichloromethane fraction</td>
<td>400</td>
<td>1.60 ± 0.22&lt;sup&gt;c&lt;/sup&gt;</td>
<td>91.27</td>
</tr>
<tr>
<td>Aqueous fraction</td>
<td>400</td>
<td>3.83 ± 0.25&lt;sup&gt;c&lt;/sup&gt;</td>
<td>79.11</td>
</tr>
<tr>
<td>Cimetidine</td>
<td>100</td>
<td>1.52 ± 0.88&lt;sup&gt;c&lt;/sup&gt;</td>
<td>91.71</td>
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</tbody>
</table>

Data were expressed as mean ± SEM. significant at ap<0.05, bp < 0.01, cp<0.001 when compared to control n = 6.

Table 2. Effect of *Homalium letestui* extract on ethanol induced ulcer.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Dose (mg/kg)</th>
<th>Ulcer indices</th>
<th>Preventive ratio (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (ethanol)</td>
<td>-</td>
<td>4.63 ± 0.33</td>
<td>-</td>
</tr>
<tr>
<td><em>Homalium letestui</em> extract p.o.</td>
<td>200</td>
<td>2.66 ± 0.33&lt;sup&gt;a&lt;/sup&gt;</td>
<td>42.54</td>
</tr>
<tr>
<td></td>
<td>400</td>
<td>2.00 ± 0.61&lt;sup&gt;b&lt;/sup&gt;</td>
<td>56.80</td>
</tr>
<tr>
<td></td>
<td>600</td>
<td>1.34 ± 0.57&lt;sup&gt;c&lt;/sup&gt;</td>
<td>71.05</td>
</tr>
<tr>
<td>Dichloromethane fraction</td>
<td>400</td>
<td>1.20 ± 0.25&lt;sup&gt;c&lt;/sup&gt;</td>
<td>74.08</td>
</tr>
<tr>
<td>Aqueous fraction</td>
<td>400</td>
<td>2.66 ± 0.22&lt;sup&gt;a&lt;/sup&gt;</td>
<td>42.54</td>
</tr>
<tr>
<td>Propranolol</td>
<td>40</td>
<td>0.82 ± 0.15&lt;sup&gt;c&lt;/sup&gt;</td>
<td>88.76</td>
</tr>
</tbody>
</table>

Data were expressed as mean ± SEM. significant at ap<0.05, bp < 0.01, cp<0.001 when compared to control n = 6.

Table 3. Effect of *Homalium letestui* extract on histamine - induced ulceration in rats.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Dose (mg/kg)</th>
<th>Ulcer indices</th>
<th>Preventive ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (Histamine)</td>
<td>100</td>
<td>16.01 ± 0.81</td>
<td>-</td>
</tr>
<tr>
<td><em>Homalium letestui</em> extract p.o.</td>
<td>200</td>
<td>12.62 ± 1.40&lt;sup&gt;a&lt;/sup&gt;</td>
<td>21.17</td>
</tr>
<tr>
<td></td>
<td>400</td>
<td>9.25 ±0.82b</td>
<td>42.22</td>
</tr>
<tr>
<td></td>
<td>600</td>
<td>3.12 ± 0.12c</td>
<td>80.51</td>
</tr>
<tr>
<td>Dichloromethane fraction</td>
<td>400</td>
<td>2.25 ±0.14c</td>
<td>85.94</td>
</tr>
<tr>
<td>Aqueous fraction</td>
<td>400</td>
<td>10.25 ± 0.66</td>
<td>35.97</td>
</tr>
<tr>
<td>Cimetidine</td>
<td>100</td>
<td>1.00 ± 0.11&lt;sup&gt;a&lt;/sup&gt;</td>
<td>94.12</td>
</tr>
</tbody>
</table>

Data were expressed as mean ± SEM. significant at *P < 0.001 when compared to control n = 6.

Table 4. Effect of *Homalium letestui* extract on reserpine induced ulceration in rats.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Dose (mg/kg)</th>
<th>Ulcer indices</th>
<th>Preventive ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (Reserpine)</td>
<td>8</td>
<td>16.22 ± 0.65</td>
<td>-</td>
</tr>
<tr>
<td><em>Homalium letestui</em> extract p.o.</td>
<td>200</td>
<td>12.56 ± 0.94&lt;sup&gt;a&lt;/sup&gt;</td>
<td>22.56</td>
</tr>
<tr>
<td></td>
<td>400</td>
<td>8.21 ± 0.53&lt;sup&gt;b&lt;/sup&gt;</td>
<td>49.38</td>
</tr>
<tr>
<td></td>
<td>600</td>
<td>3.44 ± 0.67&lt;sup&gt;b&lt;/sup&gt;</td>
<td>78.79</td>
</tr>
<tr>
<td>Dichloromethane fraction</td>
<td>400</td>
<td>2.75 ± 0.34&lt;sup&gt;b&lt;/sup&gt;</td>
<td>83.04</td>
</tr>
<tr>
<td>Aqueous fraction</td>
<td>400</td>
<td>7.56 ± 0.52&lt;sup&gt;a&lt;/sup&gt;</td>
<td>65.72</td>
</tr>
<tr>
<td>Cimetidine</td>
<td>100</td>
<td>1.10 ± 0.84&lt;sup&gt;b&lt;/sup&gt;</td>
<td>93.21</td>
</tr>
</tbody>
</table>

Data were expressed as mean ± SEM. significant at *P < 0.01; bP < 0.001 when compared to control n = 6.
5. References


Okokon, J.E., Dar, A., Choudhary, M.I. 2013b. Cellular antioxidative, anticancer and


